

Potential Role of Neurogenic Inflammatory Factors in the Pathogenesis of Vitiligo

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Background: Vitiligo is a highly complex multifactorial condition of the skin that has an unclear mechanism of pathogenesis.

Objective: This review summarizes the role of various neurogenic inflammatory factors significantly upregulated in vitiligo.

Methods: A literature review was conducted of all pertinent data regarding neuropeptides that are altered in vitiligo and their possible role in the destruction of melanocytes.

Results: The close associations between the skin, immune system, and nervous system, along with specific changes demonstrated in vitiligo patients, support a pathogenic mechanism of vitiligo that involves neuroimmunologic factors, the release of which can be governed by mental stress.

Conclusion: Neuropeptides and nerve growth factors are critical regulators of emotional response and may precipitate the onset and development of vitiligo in certain predisposed individuals. More studies are required to investigate whether a direct link exists between genetics, mental stress, and neurogenic factors in vitiligo.

Renseignements de base: Le vitiligo est une anomalie de la pigmentation de la peau ayant une étiologie multifactorielle très complexe dont le mécanisme de pathogenèse n'est pas clair.

Objectif: Ce résumé présente brièvement le rôle de divers facteurs d'inflammations neurogènes régulés à la hausse de façon significative dans le vitiligo.

Méthodes: Cet article comprend une analyse documentaire de toutes les données pertinentes sur les neuropeptides qui sont modifiés dans le vitiligo et leur rôle possible dans la destruction des mélanocytes.

Résultats: Les associations étroites entre la peau, le système immunitaire, et le système nerveux, avec des modifications particulières démontrées chez les patients atteints de vitiligo, appuient un mécanisme pathogène de vitiligo qui met en jeu des facteurs neuro-immunologiques dont la libération peut être régie par la tension mentale.

Conclusion: Les neuropeptides et des facteurs de croissance nerveuse sont des régulateurs essentiels de la réponse émotionnelle et peuvent précipiter l'apparition et le développement du vitiligo chez certaines personnes prédisposées. D'autres études sont nécessaires pour déterminer s'il existe un lien direct entre la génétique, la tension mentale, et les facteurs neurogènes du vitiligo.

VITILIGO is an acquired pigmentation disorder in which melanocytes, the principal pigment-producing cells in humans, are destroyed. With a worldwide incidence of 0.5 to 1%, vitiligo can be a highly disfiguring disorder. The resulting depigmentation can affect the retinal epithelium, the skin, the hair, and the mucous

membranes.¹⁻³ Although vitiligo is traditionally viewed as a minor disease, it has been reported to have a severe impact on the psychological well-being of the affected individuals, resulting in impaired social interactions and decreased quality of life.⁴⁻⁷ In addition to the cosmetic and psychosocial implications, there is increasing evidence of association with other autoimmune diseases, such as systemic lupus erythematosus, hypothyroidism, diabetes, and various disorders of the nervous system.⁸⁻¹⁶ Therefore, vitiligo has a major impact on the well-being of the affected individuals.

The pathogenesis of vitiligo is poorly understood. The main reason lies in the multifactorial nature of the disease, which progresses as a result of the interplay between multiple genes and environmental factors. Several prevailing pathogenic theories have dominated the mainstream literature: (1) the autoimmune theory¹⁷; (2) the neural

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hypothesis¹⁸; (3) the oxidative and redox imbalance theory (which overlaps with the neural hypothesis)¹⁹; and (4) the melanocytorrhagy or “intrinsic adhesion defect” theory.²⁰ Although each theory has its own proponents, the autoimmune hypothesis arguably has the most advocates and experimental support. However, it is likely that the pathogenesis of vitiligo involves a combination of multiple proposed mechanisms. Accumulating research has revealed intricate connections and bidirectional crosstalk between the immune system and the nervous system^{21–23} and the impact they may have on the survival and integrity of melanocytes in vitiligo skin. It is well established that immune-related cells in the skin (such as lymphocytes, macrophages, natural killer cells, and keratinocytes) express receptors for neurotransmitters and neuropeptides.^{24–28} This review highlights findings from the literature that support a pathogenic role by stress-induced neurogenic inflammatory factors in the development of vitiligo.

Evidence of Neuroendocrine Imbalance in Vitiligo

Studies have demonstrated in vitiligo lesional skin altered numbers and distribution of nerve fibers, including those that secrete neuropeptide Y (NPY) and calcitonin gene-related peptide (CGRP)^{29,30} and those that are immunoreactive for the low affinity (p75) nerve growth factor receptor (NGFr-IR).³¹ Significantly elevated levels of NPY in the plasma and tissue fluids of vitiligo patients have also been observed.³² Furthermore, it has been documented that the catecholamine neurotransmitters, such as dopamine, epinephrine, and norepinephrine, are significantly elevated in the serum and urine of vitiligo patients.^{33–37}

Role of Mental Stress in Vitiligo

There is accumulating evidence of a strong association between mental stress and the onset or progression of vitiligo. A case-control study in 2004 on children afflicted with vitiligo and psoriasis showed that the onset of vitiligo was associated with psychological factors.³⁸ Another case-control study done by Manolache and Lecca demonstrated that vitiligo patients are much more likely (odds ratio of 6.81) to encounter stressful events in their life.³⁹ Their study also revealed that patients were much more likely to experience one stressful event before the onset of vitiligo. Furthermore, it has been suggested that patients with alexithymia (deficiency in the ability to express emotions) and those with poor social support are more susceptible to vitiligo.⁴⁰

Mental stress and other psychological factors have been widely implicated in the initiation and exacerbation of

autoimmune diseases, such as systemic lupus erythematosus, rheumatoid arthritis, diabetes, and various dermatoses.⁴¹ Furthermore, it has been suggested that the stress-associated exacerbation of autoimmune or inflammatory disorders is mediated through neurotransmitters and hormones.⁴²

An important consequence of mental stress is through its effect on the secretion of catecholamines via stimulation of the hypothalamic-pituitary-adrenal (HPA) axis,^{42,43} which consists of a set of complex interactions between the hypothalamus, pituitary gland, and adrenal glands. Systemically, psychological and emotional disturbances can trigger the production and release of corticotrophin-releasing hormone (CRH) by the hypothalamus, which in turn stimulates the release of adrenocorticotrophic hormone (ACTH) by the pituitary gland. ACTH can act on the adrenal gland to produce various corticosteroids and catecholamines. More importantly, in addition to systemic effects, the HPA axis has been shown to play a crucial regulatory role in the local microenvironment of the skin, where ACTH, CRH, and CRH receptors are involved.⁴⁴ Moreover, melanocyte development and pigment production are directly regulated by local sympathetic adrenergic innervations, an increased activity of which has been shown in vitiligo lesions.^{45–47}

In addition to catecholamines, other neural-inflammatory mediators implicated in vitiligo pathogenesis, such as NPY, CGRP, nerve growth factor (NGF), and NGF receptors, are also strongly influenced by mental stress.^{48–54} NPY is known to be concentrated in the hypothalamus in the central nervous system. In the peripheral nervous system, NPY is mainly present in the sympathetic innervations and coreleased with catecholamines on nerve stimulation by mental stress.⁴⁸ In addition, NPY has been suggested to play a regulatory role in the maintenance of emotional homeostasis by either stimulating or suppressing the HPA axis depending on the plasma concentration of epinephrine and norepinephrine.^{55,56} Therefore, NPY is an important molecule involved in the HPA axis in mediating stress and anxiety.

Stress-induced upregulation of NGFs and their corresponding receptors is known to promote various neurogenic inflammatory processes by stimulating the secretion of proinflammatory neuropeptides such as CGRP and substance P (SP).^{49,54} Joachim and colleagues demonstrated that both mental stress and administration of NGF were able to significantly induce the growth of CGRP and SP-positive nerve fibers in murine skin and that the application of anti-NGF successfully abrogated the response.⁴⁹ In addition, CGRP has been shown to further stimulate the

HPA axis to produce corticosteroid hormones, such as cortisol, which in turn may induce the production of epinephrine and norepinephrine.⁵⁷ In the context of vitiligo, increased growth of CGRP-positive nerve fibers in the skin may be stimulated by the release of NGF as a result of mental stress. In addition, being one of the most potent vasodilators known,⁵⁸ CGRP may also be upregulated in response to vasoconstriction evoked by other stress-induced neuropeptides, such as NPY and catecholamines.⁵⁹

Neurogenic Inflammatory Response in Vitiligo

Numerous studies demonstrate that neurogenic factors strongly influenced by mental stress can directly and/or indirectly influence the survival and structural integrity of melanocytes. The following summarizes the evidence supporting a potential pathogenic role played by the following neurogenic factors: NPY, CGRP, catecholamines, and NGFs and NGF receptors. In general, the potential mechanisms through which these factors lead to melanocyte destruction are (1) nonspecific direct cytotoxicity to melanocytes and (2) the initiation and propagation of local and systemic immune or inflammatory reaction, including a specific adaptive immune response against melanocytes (Figure 1, Figure 2, Figure 3, and Figure 4).

Neuropeptide Y

Widely distributed in both the peripheral and the central nervous system, NPY is a neurotransmitter with diverse functions.⁶⁰ It is a stress hormone and a potent vasoconstrictor. It has been implicated in the regulation of food intake, memory, and neuroendocrine balance.^{61–65} NPY can influence the survival of melanocytes by several potential mechanisms, including indirect activation of adaptive immune response and through formation of reactive oxygen species (ROS) (see Figure 1).

There is substantial evidence of NPY playing an important role in regulating the function of cells involved in innate and adaptive immunity, such as monocytes, polymorphonuclear leukocytes (PMNs), lymphocytes, and antigen-presenting cells (APCs). In addition to enhancing the phagocytosis capabilities of APCs, such as dendritic cells (DCs), NPY has been shown to directly stimulate the production and release of ROS in PMNs and macrophages, both directly by binding to Y1 and Y5 receptors and indirectly by stimulation of the central nervous system, which has been demonstrated in mice.^{66–70} ROS may also be generated as a result of the vasoconstrictor effects exerted by NPY on endothelial cells

in the skin capillary system. Vasoconstriction activates nicotinamide adenine dinucleotide phosphate (NADPH) oxidase in endothelial cells and phagocytes, which catalyzes the formation of ROS.⁷¹

Elevated levels of oxygen radicals and peroxide have been well documented in the dermis of vitiligo patients. It has been reported that ROS can cause destruction of melanocytes via direct cytotoxic effects, deactivation of critical enzymes (ie, catalase and acetylcholinesterase), and alteration of melanocytic structural antigens or “neoantigens,” which triggers further specific adaptive immune response against melanocytes.^{72–74}

There is substantial evidence of NPY playing an important role in regulating the production of cytokines by macrophages, PMNs, and lymphocytes. This effect of NPY is exerted through its binding to various types of G-protein receptors (ARs) present on these cells. Through its stimulation of ARs, NPY can stimulate the production of proinflammatory cytokines in lymphocytes and PMNs, such as interleukin (IL)-1 β , IL-6, interferon- γ (IFN- γ), and tumor necrosis factor α (TNF- α).^{75–78} All of these cytokines were previously reported to be elevated in the serum and lesional skin of vitiligo patients.^{79,80} It is noted that NPY's effects on the secretion of proinflammatory cytokines are not universally accepted, with some reports demonstrating an increased production of IFN- γ and IL-2, skewing the immune response toward the T-helper (Th)1 pathway, whereas others reported the opposite effect, in that NPY suppresses the Th1 pathway and instead promotes the production of IL-4, shifting to a Th2-type response.^{77,81} Therefore, the precise role of NPY in the cytokine dysregulation observed in vitiligo patients remains to be further clarified.

The presence of leukocyte infiltrates—T cells, Langerhans cells (LCs), and macrophages—in the lesional and perilesional skin of vitiligo patients has been well-documented.^{82–86} NPY may play a role in this because it has been shown to have a profound impact on the trafficking of leukocytes, especially activated monocytes and T lymphocytes.^{70,78,87,88} These observations, when combined with the evidence of increased dermal NPY-positive nerve fibers and elevated levels of NPY in the plasma of vitiligo patients, strongly suggest a pathogenic role of NPY in the recruitment of immune cells in vitiligo.

Calcitonin Gene-Related Peptide

Another neural peptide secreted by primary afferent sensory neurons in the skin, CGRP,⁸⁹ also participates in the communication between the nervous and immune

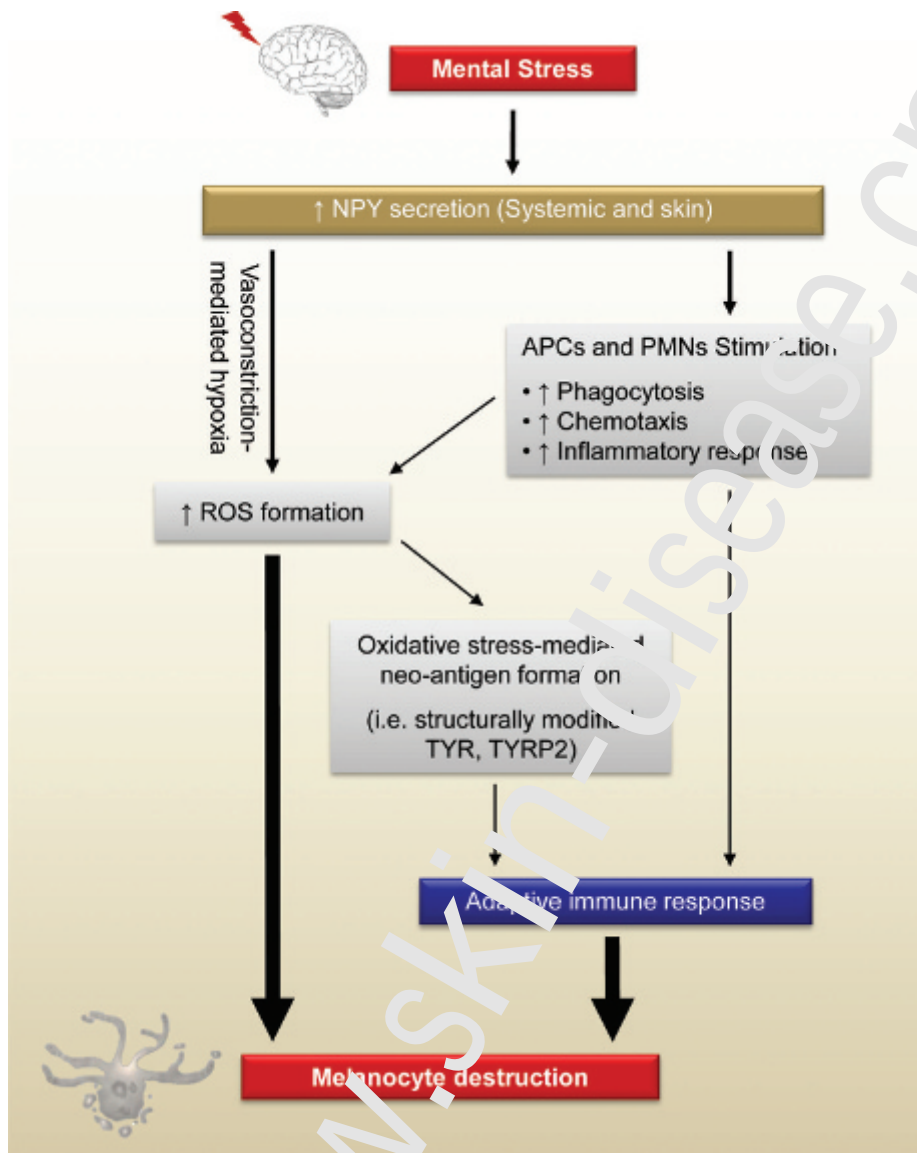


Figure 1. Proposed mechanism of neuropeptide Y-mediated melanocyte destruction. Increased release of neuropeptide Y as a result of emotional stress in genetically predisposed individuals may trigger inflammatory responses initiated by antigen-presenting cells and polymorphonuclear leukocytes. The consequences of such inflammation are increased proinflammatory cytokines and reactive oxygen species, which may cause direct melanocyte cytotoxicity. Chronic inflammatory responses would also trigger specific adaptive immune responses against melanocytes in general. APC = antigen-presenting cell; NPY = neuropeptide Y; PMN = polymorphonuclear leukocyte; ROS = reactive oxygen species; TYR = tyrosinase; TYRP2 = tyrosinase-related protein 2.

systems and potentially influences the survival of melanocytes (see Figure 2). CGRP has been shown to be associated with a variety of hypersensitivities and neurogenic diseases, such as the common migraine, temporomandibular joint disorder, rhinosinusitis, and atopic dermatitis.^{90–92} More interestingly, many immune and nonimmune cells of the skin, such as mast cells, neutrophils, LCs, lymphocytes, Schwann cells, keratinocytes, and fibroblasts, express CGRP receptors, which, when stimulated, could result in inflammatory responses, possibly through the activation of the mitogen-activated protein (MAP) kinase signaling pathways.^{78,93,94} For example, Levite and colleagues demonstrated in several studies that in addition to enhancing the adhesion capability of lymphocytes to the extracellular matrix, CGRP, along with other neuropeptides, can

stimulate the secretion of various proinflammatory cytokines by naive and mature Th1 and Th2 antigen-specific T cells, suggesting that CGRP may augment both Th1 and Th2 adaptive immune activities.^{78,94}

The effect that CGRP has on traditionally nonimmune cells can be best demonstrated in a study done by Vause and Durham on peripheral glial cells and Schwann cells, which, when cultured with the CGRP, produced drastically elevated levels of proinflammatory cytokines such as IL-1, 6, IFN- γ , and TNF- α .⁹⁵ Furthermore, keratinocytes in the presence of CGRP exhibited an increased proliferation rate and production of IL-8, which is a potent chemoattractant for macrophages and neutrophils.⁹⁶

Given that LCs are closely associated with CGRP-positive nerve fibers in the skin,⁹⁷ and there is evidence

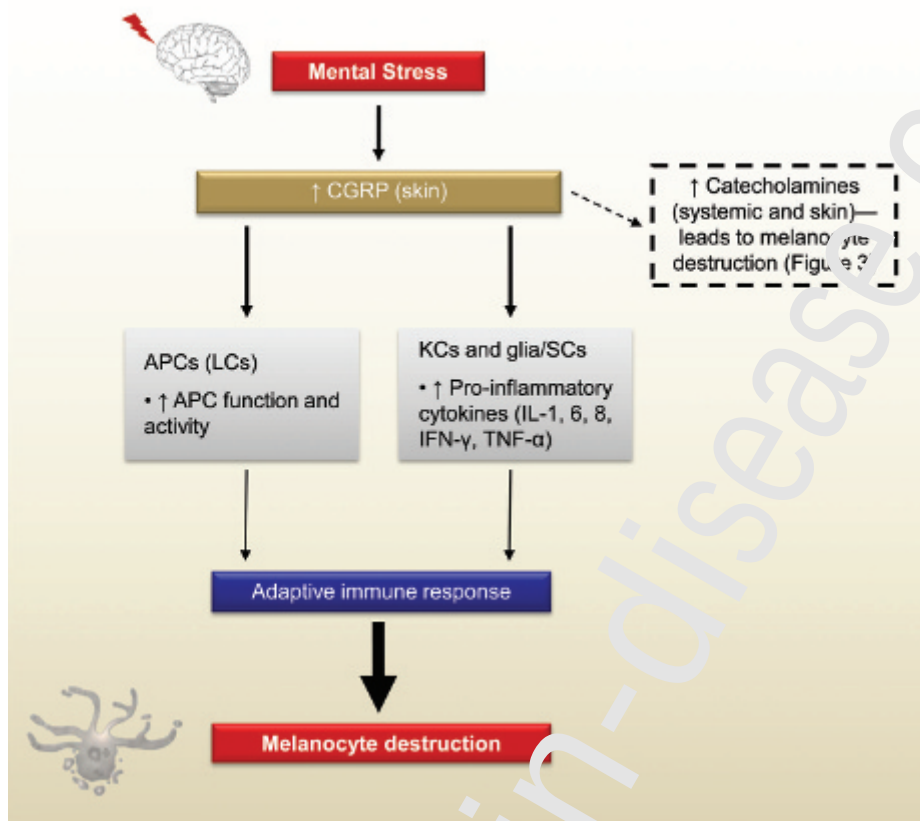


Figure 2. Proposed mechanism of calcitonin gene-related peptide (CGRP)-mediated melanocyte destruction. Emotional stress and anxiety can trigger increased production and release of CGRPs from sympathetic nerve endings in the skin. Chronic release of CGRP may trigger local and systemic inflammation by direct activation of keratinocytes, Langerhans cells, and glial cells in the skin. In addition, CGRP can induce the release of catecholamines through stimulation of the hypothalamus-pituitary-adrenal axis. APC = antigen-presenting cell; IFN- γ = interferon- γ ; IL = interleukin; KC = keratinocyte; LC = Langerhans cell; SC = Schwann cell; TNF- α = tumor necrosis factor α .

that CGRP augments the antigen-presenting ability of LCs,⁹⁸ it is possible that the coordination and stimulation of LCs by CGRP represent an early step leading to the melanocyte-specific adaptive immune responses observed in vitiligo. Another potential mechanism through which CGRP may influence melanocyte survival is via its ability to stimulate catecholamine secretion in vitiligo⁵⁷ (see Figure 3).

Catecholamines

The death of melanocytes in vitiligo has been attributed to increased oxidative stress caused by the accumulation of sympathetic catecholamine neurotransmitters and their metabolites (see Figure 3). For example, multiple studies have demonstrated increased urinary and serum levels of dopamine, epinephrine, and norepinephrine in vitiligo patients compared to controls.^{33–37} In addition, the oxidation products of epinephrine and norepinephrine, such as homovanillic acid (HVA) and vanillylmandelic acid (VMA), were also found in significantly elevated levels in the urine of vitiligo patients.^{33,35} The main consequence of catecholamine accumulation may be the

production of peroxide and toxic oxygen radicals as a result of (1) metabolic breakdown of catecholamines by monoamine oxidases, which has been observed to be upregulated in vitiligo lesional skin,^{36,99} and (2) vasoconstriction and subsequent hypoxia induced by norepinephrine, which has also been shown to activate monoamine oxidases, possibly as a regulatory mechanism for its own degradation.¹⁰⁰

Recent studies have shown that the monoamine neurotransmitters, especially epinephrine and norepinephrine, influence immune responses primarily via direct binding to high-affinity α and β adrenoreceptors that are present on most leukocytes. It has been demonstrated that epinephrine-treated DCs can elicit the production of various cytokines by T lymphocytes, such as IL-4, IL-10, IL-12, and, most notably, IL-17, a finding that implicated epinephrine in skewing the immune responses toward either the Th2 or the Th17 pathway.¹⁰¹ The Th17 pathway has been implicated in the initiation and progression of autoimmune diseases.^{102–107} IL-17 is a potent inducer of other proinflammatory cytokines such as IL-1, IL-6, and TNF- α .^{108–111} Moreover, a significantly elevated level of IL-17 has been observed in both the serum and the tissue fluid of vitiligo patients.¹¹²

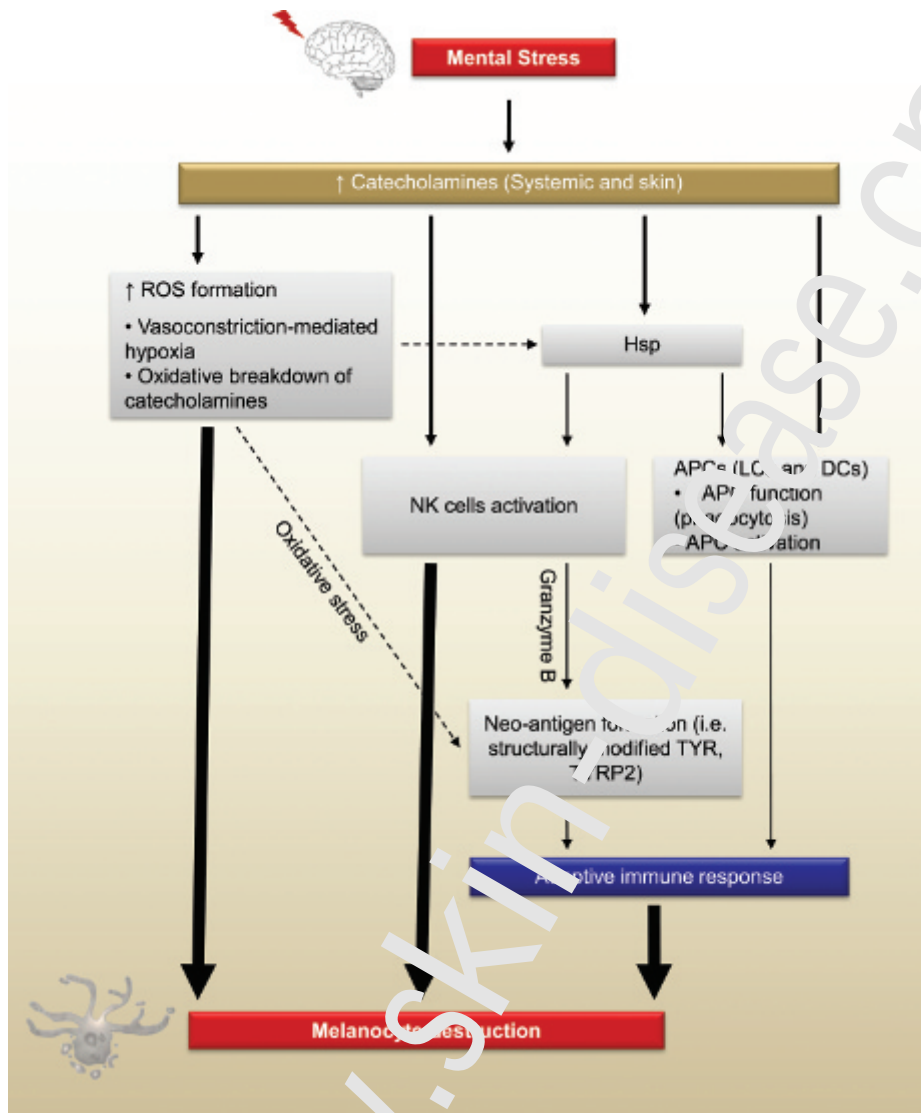


Figure 3. Proposed mechanism of catecholamine-mediated melanocyte destruction. Emotional stress can directly stimulate the hypothalamus-pituitary-adrenal axis to release catecholamines from the adrenal gland as well as adrenergic nerve endings in the skin. The increased levels of catecholamines and their metabolic products may mediate immune responses via several pathways: (1) direct stimulation and activation of natural killer cells, which may cause direct cytotoxicity of melanocytes through granzyme B-mediated apoptosis and possible formation of neoantigens; (2) stimulate the release of heat shock proteins by melanocytes, which may then trigger an inflammatory response involving the recruitment and activation of antigen-presenting cells followed by the development of adaptive immune responses; (3) direct activation of antigen-presenting cells such as dendritic cells and Langerhans cells; and (4) formation and accumulation of reactive oxygen species, which, besides causing direct melanocyte destruction, would also trigger an inflammatory cascade, leading to the activation of adaptive immunity. APC = antigen-presenting cell; DC = dendritic cell; Hsp = heat shock protein; LC = Langerhans cell; NK = natural killer; ROS = reactive oxygen species; TYR = tyrosinase; TYRP2 = tyrosinase-related protein 2.

A study by Dimitroff and colleagues showed that epinephrine can selectively recruit and mobilize cytotoxic leukocytes, such as CD8⁺ T cells, CD3⁺CD56⁺ natural killer T (NKT)-like cells, and natural killer cells.¹¹³ Both cytotoxic CD8⁺ T cells and natural killer cells have been implicated as playing a direct role in the destruction of melanocytes in vitiligo as they were found in a significantly increased number in either the blood or the lesional skin of vitiligo patients.^{82,114}

There is evidence that norepinephrine may also play a prominent role during the immune response, such as in augmenting antigen uptake by macrophages and stimulating natural killer cells. Norepinephrine has been shown to induce increased endocytosis by DCs via α_2 -adrenoreceptor stimulation and activation of the PI3K and ERK1/2 intracellular signaling pathways.¹¹⁵ Norepinephrine was

also able to increase the cytotoxicity of natural killer cells, possibly by direct stimulation via α -adrenoreceptors.^{116,117}

Catecholamines have previously been demonstrated to induce intracellular heat shock protein (Hsp)72 in tissues via direct stimulation of α_2 -adrenergic receptors.^{118–121} Hsps are stress-inducible proteins present in all cells and are crucial in the induction of innate immunity, especially in response against cancer.^{122–124} Hsps are known to enhance both innate and adaptive immunologic responses by stimulating the proliferation of DCs and cytotoxic T cells as well as the release of proinflammatory cytokines via a calcium-dependent pathway.^{125–131} The Hsp70 family is also known to enhance the cytotoxic activity of natural killer cells^{122,132} and induce the secretion of proinflammatory cytokines and proteases, such as IFN- γ and granzyme B, which, besides direct cytotoxic effects, has also been

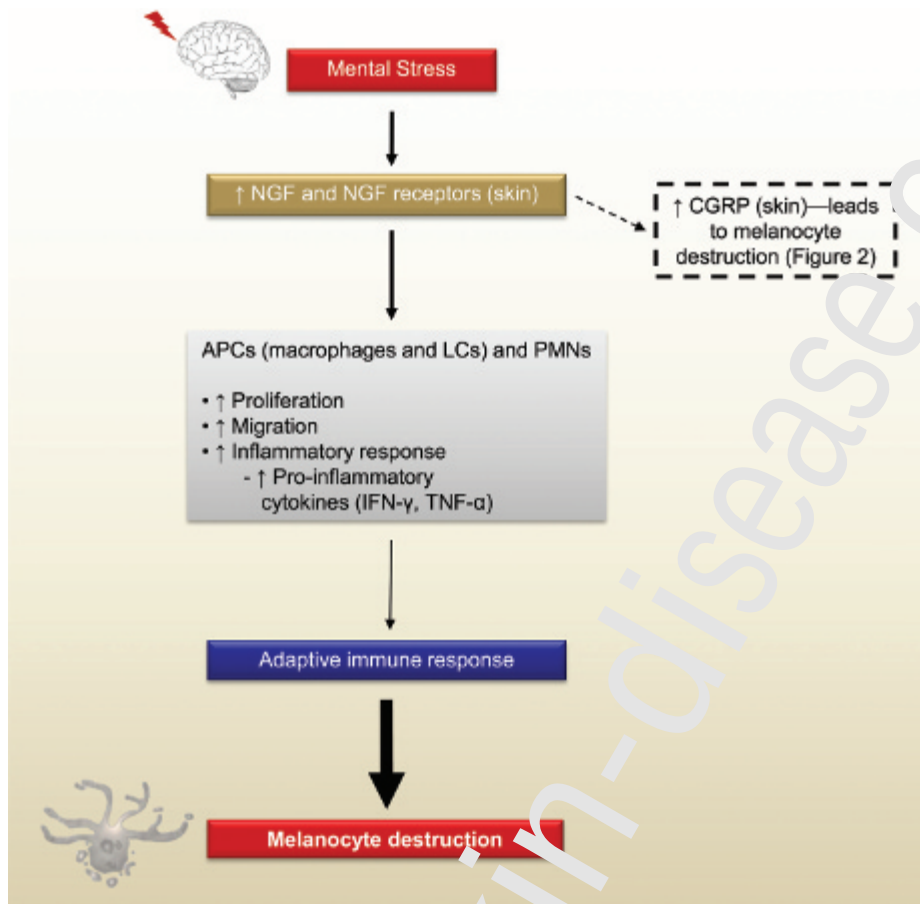


Figure 4. Proposed mechanism of nerve growth factor (NGF)-mediated melanocyte destruction. NGFs are suggested to be important mediators of neurogenic inflammation in many autoimmune diseases. NGFs have been demonstrated to play a critical role in the development of both cell-mediated and humoral adaptive immunity and are especially important in the maintenance of immunologic memory. In addition to the direct effect on the activity of immune cells, NGFs are also potent stimulants for the growth of peptidergic nerve endings in the skin and thus are able to induce the release of additional neuropeptides, which include CGRP. APC = antigen-presenting cell; CGRP = calcitonin gene-related peptide; IFN- γ = interferon- γ ; LC = Langerhans cell; PMN = polymorphonuclear leukocyte; TNF- α = tumor necrosis factor α .

implicated in the generation of “neointensities” in autoimmune diseases.^{74,133} Recent studies have shown that Hsps may play a role in the pathogenesis of vitiligo. Histologic studies have revealed consistent differential expression of Hsp70 in the lesion and perilesional skin of vitiligo patients.¹³⁴ Denman and colleagues recently demonstrated in a mouse model of autoimmune vitiligo that gene gun vaccination of expression vectors containing tyrosinase-related protein 2 (Trp-2) along with Hsp70 significantly accelerated the repigmentation process compared to control (Trp-2 only) vectors.¹³⁵ Hsps are also expressed in times of hypoxia and oxidative stress to augment the activity of antioxidant enzymes to protect cell viability.^{136–139} In the skin, Hsp70 can be readily induced in fibroblasts with peroxides.^{140,141} Interestingly, Kroll and colleagues demonstrated in their study that oxidative stress (in the form of the chemical 4-tertiary butyl phenol) was able to induce the secretion of Hsp70 by melanocytes cultured in vitro, which in turn induced the upregulation of membrane tumor necrosis factor-related apoptosis-inducing ligand (TRAIL) expression and subsequently enhanced DC-mediated killing of melanocytes.⁸⁶

This suggests that in the context of vitiligo, the accumulation of catecholamines will not only directly induce the expression of Hsps in melanocytes via adrenoreceptor interaction¹⁴² but may also stimulate the release of Hsps as a result of oxidative stress secondary to vasoconstriction and metabolic breakdown. The upregulation of Hsps would in turn augment the innate and adaptive immune responses against melanocytes.

NGFs and Their Receptors

Neurotrophins of the NGF family are essential trophic factors secreted by the hypothalamus for the development and maintenance of neurons and other cells derived from neural origin.^{143,144} It has been shown that NGF binds to two types of receptors distinguished from each other based on their specificity and affinity for particular neurotrophins: the trkA tyrosine kinase receptor and the p75 NGF receptor.^{145,146} The trkA receptor has high affinity and is very specific for certain neurotrophins, whereas the p75 NGFr binds to virtually all members of the NGF family with lower but equal affinity. In general, the effect of NGF

on target tissues depends on the expression level of NGF receptors (both trkA and p75). In the skin, NGF is crucial for the maintenance of sympathetic nerve fibers.¹⁴⁷ This effect is evident in the skin of vitiligo patients, where increased innervations of the aforementioned nerve fibers (NPY, CGRP, p75 NGFr-IR) have been observed. The significant upregulation in the expression of p75 NGFr in vitiligo skin also implies the hyperresponsiveness of the dermal and epidermal environment to NGF in general.

There is increasing evidence that attributes NGF to the role of an important messenger between the nervous system and the immune system. Studies have demonstrated that NGFs are produced by most, if not all, of the major players in the immune system—monocytes/macrophages, neutrophils, granulocytes, and lymphocytes—all of which also express both types of NGF receptors.^{148–157} Therefore, NGF is able to influence their proliferation, differentiation, and other functional aspects, such as migration through vascular endothelium during inflammatory responses, in an autocrine and paracrine manner.^{147,150,152,155} These activities imply that NGF and its receptors have a potential role in the destruction of melanocytes in vitiligo (see Figure 4).

NGF and the corresponding trkA and p75 receptors have been suggested to stimulate the proliferation and differentiation of T and B lymphocytes.^{150,155,156} NGF was also suggested to be involved in the development and maintenance of immunologic memory in adaptive immunity.^{150,155} Given the suggested hyperresponsiveness of vitiligo skin to NGF and the presence of cytotoxic T lymphocytes and autoantibodies in vitiligo patients against melanocyte-associated markers such as tyrosinase and tyrosinate-related proteins,^{158,161} it is not unreasonable to assume that the NGF receptors on immune cells of vitiligo patients may also be upregulated and that NGF may play a very important role in the propagation and potentiation of both cell-mediated and humoral response against melanocytes.

Two of the major proinflammatory cytokines that have been observed in vitiligo patients, IFN- γ and TNF- α , could be readily induced by NGF in macrophages, mast cells, and eosinophils, which in turn may produce more NGF along with inflammatory cytokines.^{153,162–164} Furthermore, NGF has been implicated in various autoimmune diseases and allergic conditions, such as systemic lupus erythematosus, psoriasis, rheumatoid arthritis, asthma, and urticaria, where serum levels of NGF were significantly increased in patients.^{136,139,140} Recently, NGF has been demonstrated to exacerbate inflammation in a mouse model of atopic allergic dermatitis, whereas administration of anti-NGF-

neutralizing antibodies substantially alleviated symptoms, including the suppression of TNF- α .¹⁴¹ As some of these diseases are also associated with vitiligo, it may be worthwhile to further investigate the role of NGF as a mediator in the destruction of melanocytes in the future.

Implications on Therapy

Presently, treatment options for vitiligo patients typically involve the direct modulation of immune responses to melanocytes through local immunosuppression with topical corticosteroids, such as clobetasol propionate and betamethasone valerate, and calcineurin inhibitors, such as cyclosporine, pimecrolimus, and tacrolimus,^{165,166} which have been shown to stimulate the production of IL-10,¹⁶⁷ an antiinflammatory Th2 cytokine known to counteract excessive immunity in contact dermatitis and Crohn disease.^{168,169} Systemic treatment such as oral dexamethasone is also used, albeit with common side effects, including weight gain, acne, and menstrual irregularities in women.¹⁷⁰ Phototherapy such as psoralen in combination with ultraviolet A (PUVA) and narrow-band ultraviolet B (NB-UVB) has also been the mainstream treatment option for vitiligo.

To date, the findings of neural-inflammatory interactions in vitiligo have not been translated into therapeutic advances for patients, whereas neuropeptide receptor antagonists are commonly used in internal medicine (ie, gastroenterology and cardiology). For example, α and β -adrenoreceptor antagonists are used extensively in myocardial infarction and hypertension due to their ability to reverse the effects of catecholamines by acting on adrenergic receptors on endothelial cells. There is also evidence of inflammation suppression by α -adrenoreceptor antagonists, which were shown to downregulate the production of proinflammatory cytokines by lamina propria mononuclear cells and subsequently alleviated acute murine colitis.¹⁷¹ In addition, α -blockers and β -blockers are also used to treat anxiety and panic disorders by directly inhibiting the release of epinephrine and norepinephrine.^{172,173} Therefore, adrenoreceptor antagonists may be potential therapeutic candidates for vitiligo by acting as neuroinflammatory modulators and vasodilators to counter the oxidative stress generated secondary to the vasoconstrictor effect of neuropeptides on endothelial cells. Adrenoreceptor antagonists may potentially complement the immunosuppressants currently prescribed in treating vitiligo, such as topical tacrolimus, which, when used alone, has actually been demonstrated to induce the production and release of neuropeptides in the skin.¹⁷⁴

Given the association between emotional stress and vitiligo, management and treatment strategies traditionally employed in the field of psychiatry have also demonstrated some success in vitiligo. Antidepressants and antipsychotropic medications were effective in controlling the disease progression when used alone or in conjunction with other treatment options,¹⁷⁵ a finding that has yet to be replicated by others. In addition, there is some preliminary evidence that cognitive behavioral therapy, stress management strategies, and other psychological educational programs have been effective in ameliorating vitiligo severity.^{5,176}

Conclusion

Given that neuropeptides and hormones are critical regulators of emotional response and only a small fraction of individuals exposed to mentally stressful situations develop vitiligo, additional factors must be involved in vitiligo pathogenesis. Of major importance is genetic predisposition, as revealed in multiple studies, all of which identified genetic markers that are strongly associated with the development of vitiligo.^{177–187} It is worth noting that most of these implicated genetic differences in vitiligo patients are located in genes or chromosomal regions that regulate the innate and, to a lesser extent, adaptive immunity and inflammation, such as *NALP1* and certain major histocompatibility complex alleles. On the other hand, there have been no experimental data linking these genetic findings with mental stress and neurogenic peptides, although most of the implications of genetic elements have yet to be characterized in terms of their precise physiologic functions. More experimental investigations are warranted to fully understand the role of neurogenic mediators in vitiligo and their potential implications in the development of therapy.

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References

1. Abu Tahir M, Pramod K, Ansari SH, et al. Current remedies for vitiligo. *Autoimmun Rev* 2010;9:516–20, doi:[10.1016/j.autrev.2010.02.013](https://doi.org/10.1016/j.autrev.2010.02.013).
2. Grimes PE. New insights and new therapies in vitiligo. *JAMA* 2005;293:730–5, doi:[10.1001/jama.293.6.730](https://doi.org/10.1001/jama.293.6.730).
3. Whitton ME, Ashcroft DM, Gonzalez U. Therapeutic interventions for vitiligo. *J Am Acad Dermatol* 2008;59:713–7, doi:[10.1016/j.jaad.2008.06.023](https://doi.org/10.1016/j.jaad.2008.06.023).
4. Porter JR, Beuf AH. Racial variation in reaction to physical stigma: a study of degree of disturbance by vitiligo among black and white patients. *Health Soc Behav* 1991;32:192–204, doi:[10.2307/2137152](https://doi.org/10.2307/2137152).
5. Thompson AR, Kenner G, Smith JA. Living with vitiligo: dealing with difference. *Am J Health Psychol* 2002;7:213–25, doi:[10.1348/135910702169477](https://doi.org/10.1348/135910702169477).
6. Porter JR, Beuf AH, Jorner A, et al. Psychosocial effect of vitiligo: a comparison of vitiligo patients with “normal” control subjects, with psoriasis patients, and with patients with other pigmentary disorders. *J Am Acad Dermatol* 1986;15:220–4, doi:[10.1016/S0190-9622\(86\)70160-6](https://doi.org/10.1016/S0190-9622(86)70160-6).
7. Kenner G, Al'Abadie M. Psychologic effects of vitiligo: a critical incident analysis. *J Am Acad Dermatol* 1996;35:895–8, doi:[10.1016/S0190-9622\(96\)90112-7](https://doi.org/10.1016/S0190-9622(96)90112-7).
8. Boekholt K, Newby PR, Simmonds MJ, et al. Prevalence and relative risk of other autoimmune diseases in subjects with autoimmune thyroid disease. *Am J Med* 2010;123:183 e181–9, doi:[10.1016/j.amjmed.2009.06.030](https://doi.org/10.1016/j.amjmed.2009.06.030).
9. Terenzi G, Kountouras J, Koutlas E, et al. Familial prevalence of autoimmune disorders in multiple sclerosis in northern Greece. *Mult Scler* 2010;16:1091–101, doi:[10.1177/1352458510375708](https://doi.org/10.1177/1352458510375708).
10. Kocer B, Nazliel B, Oztas M, et al. Vitiligo and multiple sclerosis in a patient treated with interferon beta-1a: a case report. *Eur J Neurol* 2009;16:e78–9, doi:[10.1111/j.1468-1331.2009.02563.x](https://doi.org/10.1111/j.1468-1331.2009.02563.x).
11. Ramagopalan SV, Dymant DA, Valdar W, et al. Autoimmune disease in families with multiple sclerosis: a population-based study. *Lancet Neurol* 2007;6:604–10, doi:[10.1016/S1474-4422\(07\)70132-1](https://doi.org/10.1016/S1474-4422(07)70132-1).
12. Rashtak S, Pittelkow MR. Skin involvement in systemic autoimmune diseases. *Curr Dir Autoimmun* 2008;10:344–58, doi:[10.1159/000131754](https://doi.org/10.1159/000131754).
13. Sabate M, Bosch A, Pedros C, et al. Vitiligo associated with tolcapone and levodopa in a patient with Parkinson's disease. *Ann Pharmacother* 1999;33:1228–9, doi:[10.1345/aph.19090](https://doi.org/10.1345/aph.19090).
14. Varoglu AO, Kocaturk I, Tatar A. Prothrombin G20210A mutation, hypogonadotropic hypogonadism, and generalized vitiligo-related ischemic stroke in a young adult. *Int J Neurosci* 2010;120:451–3, doi:[10.3109/00207451003797702](https://doi.org/10.3109/00207451003797702).
15. Nikiforidis GC, Tsambaos DG, Karamitsos DS, et al. Abnormalities of the auditory brainstem response in vitiligo. *Scand Audiol* 1993;22:97–100, doi:[10.3109/01050399309046024](https://doi.org/10.3109/01050399309046024).
16. Yacubian EM, Rosemberg S, Garrido Neto TL, et al. Rasmussen encephalitis associated with segmental vitiligo of the scalp: clinicopathologic report. *J Child Neurol* 2001;16:374–7, doi:[10.1177/088307380101600513](https://doi.org/10.1177/088307380101600513).
17. Rezaei N, Gavalas NG, Weetman AP, et al. Autoimmunity as an aetiological factor in vitiligo. *J Eur Acad Dermatol Venereol* 2007;21:865–76, doi:[10.1111/j.1468-3083.2007.02228.x](https://doi.org/10.1111/j.1468-3083.2007.02228.x).
18. Yu HS. Melanocyte destruction and repigmentation in vitiligo: a model for nerve cell damage and regrowth. *J Biomed Sci* 2002;9:564–73, doi:[10.1007/BF02254984](https://doi.org/10.1007/BF02254984).
19. Dell'anna ML, Picardo M. A review and a new hypothesis for non-immunological pathogenetic mechanisms in vitiligo. *Pigment Cell Res* 2006;19:406–11, doi:[10.1111/j.1600-0749.2006.00333.x](https://doi.org/10.1111/j.1600-0749.2006.00333.x).
20. Gauthier Y, Cario Andre M, Taieb A. A critical appraisal of vitiligo etiologic theories. Is melanocyte loss a melanocytorrhagy?

- Pigment Cell Res 2003;16:322–32, doi:[10.1034/j.1600-0749.2003.00070.x](https://doi.org/10.1034/j.1600-0749.2003.00070.x).
21. Elenkov IJ, Wilder RL, Chrousos GP, et al. The sympathetic nerve—an integrative interface between two supersystems: the brain and the immune system. *Pharmacol Rev* 2000;52:595–638.
22. Downing JE, Miyan JA. Neural immunoregulation: emerging roles for nerves in immune homeostasis and disease. *Immunol Today* 2000;21:281–9, doi:[10.1016/S0167-5699\(00\)01635-2](https://doi.org/10.1016/S0167-5699(00)01635-2).
23. Straub RH, Westermann J, Scholmerich J, et al. Dialogue between the CNS and the immune system in lymphoid organs. *Immunol Today* 1998;19:409–13, doi:[10.1016/S0167-5699\(98\)01297-3](https://doi.org/10.1016/S0167-5699(98)01297-3).
24. Blalock JE, Weigent DA. Pituitary control of immune cells. *Immunol Today* 1994;15:39, doi:[10.1016/0167-5699\(94\)90205-4](https://doi.org/10.1016/0167-5699(94)90205-4).
25. Besedovsky HO, del Rey A. Immune-neuro-endocrine interactions: facts and hypotheses. *Endocr Rev* 1996;17:64–102.
26. Madden KS, Felten DL. Experimental basis for neural-immune interactions. *Physiol Rev* 1995;75:77–106.
27. Yu XJ, Li CY, Xu YH, et al. Calcitonin gene-related peptide increases proliferation of human HaCaT keratinocytes by activation of MAP kinases. *Cell Biol Int* 2009;33:144–8, doi:[10.1016/j.cellbi.2009.07.003](https://doi.org/10.1016/j.cellbi.2009.07.003).
28. Gruber JV, Holtz R. Examining communication between ultraviolet (UV)-damaged cutaneous nerve cells and epidermal keratinocytes in vitro. *Toxicol Ind Health* 2009;25:25–30, doi:[10.1177/0748233709103412](https://doi.org/10.1177/0748233709103412).
29. Al'Abadie MS, Senior HJ, Bleehen SS, et al. Neuropeptide and neuronal marker studies in vitiligo. *Br J Dermatol* 1994;131:160–5, doi:[10.1111/j.1365-2133.1994.tb08486.x](https://doi.org/10.1111/j.1365-2133.1994.tb08486.x).
30. Hristakieva E, Lazarova R, Lazarov N, et al. Markers for vitiligo related neuropeptides in human skin nerve fibers. *Acta Med Croatica* 2000;54:53–7.
31. Liu PY, Bondesson L, Lontz W, et al. The occurrence of cutaneous nerve endings and neuropeptides in vitiligo vulgaris: a case-control study. *Arch Dermatol Res* 1996;288:470–5, doi:[10.1007/BF02505276](https://doi.org/10.1007/BF02505276).
32. Tu C, Zhao D, Lin X. Levels of neuropeptide Y in the plasma and skin tissue fluids of patients with vitiligo. *J Dermatol Sci* 2001;27:178–2, doi:[10.1016/S0923-1811\(01\)00134-7](https://doi.org/10.1016/S0923-1811(01)00134-7).
33. Morrone A, Picardo M, de Luca C, et al. Catecholamines and vitiligo. *Pigment Cell Res* 1992;5:65–9, doi:[10.1111/j.1600-0749.1992.tb00003.x](https://doi.org/10.1111/j.1600-0749.1992.tb00003.x).
34. Cucchi ML, Frattini P, Santagostino G, et al. Catecholamines increase in the urine of non-segmental vitiligo especially during its active phase. *Pigment Cell Res* 2003;16:111–6, doi:[10.1034/j.1600-0749.2003.00015.x](https://doi.org/10.1034/j.1600-0749.2003.00015.x).
35. Cucchi ML, Frattini P, Santagostino G, et al. Higher plasma catecholamine and metabolite levels in the early phase of nonsegmental vitiligo. *Pigment Cell Res* 2000;13:28–32, doi:[10.1034/j.1600-0749.2000.130106.x](https://doi.org/10.1034/j.1600-0749.2000.130106.x).
36. Schallreuter KU, Wood JM, Pittelkow MR, et al. Increased monoamine oxidase A activity in the epidermis of patients with vitiligo. *Arch Dermatol Res* 1996;288:14–8, doi:[10.1007/BF02505037](https://doi.org/10.1007/BF02505037).
37. Chu CY, Liu YL, Chiu HC, et al. Dopamine-induced apoptosis in human melanocytes involves generation of reactive oxygen species. *Br J Dermatol* 2006;154:1071–9, doi:[10.1111/j.1365-2133.2006.07293.x](https://doi.org/10.1111/j.1365-2133.2006.07293.x).
38. Barisic-Drusko V, Rucevic I. Trigger factors in childhood psoriasis and vitiligo. *Coll Antropol* 2004;28:277–85.
39. Manolache L, Banea V. Stress in patients with alopecia areata and vitiligo. *J Eur Acad Dermatol Venerol* 2007;21:921–8, doi:[10.1111/j.1468-3083.2006.02106.x](https://doi.org/10.1111/j.1468-3083.2006.02106.x).
40. Picardi A, Pasquini P, Cazzurza MS, et al. Stressful life events, social support, attachment security and alexithymia in vitiligo. A case-control study. *Psychother Psychosom* 2003;72:150–8, doi:[10.1159/000069711](https://doi.org/10.1159/000069711).
41. Stojanovich L. Stress and autoimmunity. *Autoimmun Rev* 2010;9:A271–6, doi:[10.1016/j.autrev.2009.11.014](https://doi.org/10.1016/j.autrev.2009.11.014).
42. Tolis G, Stefanis C. Depression: biological and neuroendocrine aspects. *J Hum Biol Pharmacother* 1983;37:316–22.
43. Stokes PL, Stokes CR. The hypothalamic-pituitary-adrenocortical axis in major depression. *Endocrinol Metab Clin North Am* 1988;1:171–19.
44. Sominski A, Wortsman J, Tuckey RC, et al. Differential expression of HPA axis homolog in the skin. *Mol Cell Endocrinol* 2007;265-6:143–9, doi:[10.1016/j.mce.2006.12.012](https://doi.org/10.1016/j.mce.2006.12.012).
45. McGuire J. Adrenergic control of melanocytes. *Arch Dermatol* 1970;101:173–80, doi:[10.1001/archderm.1970.04000020043007](https://doi.org/10.1001/archderm.1970.04000020043007).
46. Bir LS, Aktan S. Sympathetic skin response in psoriasis and vitiligo. *J Auton Nerv Syst* 1999;77:68–71, doi:[10.1016/S0165-1838\(99\)00030-2](https://doi.org/10.1016/S0165-1838(99)00030-2).
47. Schallreuter KU, Wood JM, Pittelkow MR, et al. Increased in vitro expression of beta 2-adrenoceptors in differentiating lesional keratinocytes of vitiligo patients. *Arch Dermatol Res* 1993;285:216–20, doi:[10.1007/BF00372012](https://doi.org/10.1007/BF00372012).
48. Ekblad E, Edvinsson L, Wahlestedt C, et al. Neuropeptide Y co-exists and co-operates with noradrenaline in perivascular nerve fibers. *Regul Pept* 1984;8:225–35, doi:[10.1016/0167-0115\(84\)90064-8](https://doi.org/10.1016/0167-0115(84)90064-8).
49. Joachim RA, Kuhlmei A, Dinh QT, et al. Neuronal plasticity of the “brain-skin connection”: stress-triggered up-regulation of neuropeptides in dorsal root ganglia and skin via nerve growth factor-dependent pathways. *J Mol Med* 2007;85:1369–78, doi:[10.1007/s00109-007-0236-8](https://doi.org/10.1007/s00109-007-0236-8).
50. Aloe L, Bracci-Laudiero L, Alleva E, et al. Emotional stress induced by parachute jumping enhances blood nerve growth factor levels and the distribution of nerve growth factor receptors in lymphocytes. *Proc Natl Acad Sci U S A* 1994;91:10440–4, doi:[10.1073/pnas.91.22.10440](https://doi.org/10.1073/pnas.91.22.10440).
51. Tometten M, Klapp BF, Joachim R, et al. Nerve growth factor and its functional receptor TrkA are up-regulated in murine decidual tissue of stress-triggered and substance P-mediated abortion. *Am J Reprod Immunol* 2004;51:86–93, doi:[10.1046/j.8755-8920.2003.00123.x](https://doi.org/10.1046/j.8755-8920.2003.00123.x).
52. Singh LK, Pang X, Alexacos N, et al. Acute immobilization stress triggers skin mast cell degranulation via corticotropin releasing hormone, neurotensin, and substance P: a link to neurogenic skin disorders. *Brain Behav Immun* 1999;13:225–39, doi:[10.1006/brbi.1998.0541](https://doi.org/10.1006/brbi.1998.0541).
53. Peters EM, Kuhlmei A, Tobin DJ, et al. Stress exposure modulates peptidergic innervation and degranulates mast cells in murine skin. *Brain Behav Immun* 2005;19:252–62, doi:[10.1016/j.bbi.2004.08.005](https://doi.org/10.1016/j.bbi.2004.08.005).
54. Peters EM, Handjiski B, Kuhlmei A, et al. Neurogenic inflammation in stress-induced termination of murine hair growth is promoted by nerve growth factor. *Am J Pathol* 2004;165:259–71, doi:[10.1016/S0002-9440\(10\)63294-4](https://doi.org/10.1016/S0002-9440(10)63294-4).

55. Haas DA, George SR. Neuropeptide Y-induced effects on hypothalamic corticotropin-releasing factor content and release are dependent on noradrenergic/adrenergic neurotransmission. *Brain Res* 1989;498:333–8, doi:[10.1016/0006-8993\(89\)91112-8](#).
56. Thorsell A. Brain neuropeptide Y and corticotropin-releasing hormone in mediating stress and anxiety. *Exp Biol Med* (Maywood) 2010;235:1163–7, doi:[10.1258/ebm.2010.009331](#).
57. Dhillon WS, Small CJ, Jethwa PH, et al. Paraventricular nucleus administration of calcitonin gene-related peptide inhibits food intake and stimulates the hypothalamo-pituitary-adrenal axis. *Endocrinology* 2003;144:1420–5, doi:[10.1210/en.2002-220902](#).
58. Brain SD, Williams TJ, Tippins JR, et al. Calcitonin gene-related peptide is a potent vasodilator. *Nature* 1985;313:54–6, doi:[10.1038/313054a0](#).
59. Wahlestedt C, Edvinsson L, Ekblad E, et al. Neuropeptide Y potentiates noradrenaline-evoked vasoconstriction: mode of action. *J Pharmacol Exp Ther* 1985;234:735–41.
60. Thorsell A, Heilig M. Diverse functions of neuropeptide Y revealed using genetically modified animals. *Neuropeptides* 2002;36:182–93, doi:[10.1054/npep.2002.0897](#).
61. Hanson ES, Dallman MF. Neuropeptide Y (NPY) may integrate responses of hypothalamic feeding systems and the hypothalamo-pituitary-adrenal axis. *J Neuroendocrinol* 1995;7:27–9, doi:[10.1111/j.1365-2826.1995.tb00757.x](#).
62. Kuo LE, Kitlinska JB, Tilan JU, et al. Neuropeptide Y acts directly in the periphery on fat tissue and mediates stress-induced obesity and metabolic syndrome. *Nat Med* 2007;13:803–11, doi:[10.1038/nm1611](#).
63. Yehuda R, Brand S, Yang RK. Plasma neuropeptide Y concentrations in combat exposed veterans: relationships to trauma exposure, recovery from PTSD, and coping. *Biol Psychiatry* 2006;59:660–3, doi:[10.1016/j.biopsych.2005.08.027](#).
64. White BD, Dean RG, Edwards GL, et al. Type II corticosteroid receptor stimulation increases NPY gene expression in basomedial hypothalamus of rats. *Am J Physiol* 1995;268:R1123–9.
65. Colmers WF, El Bahh B. Neuropeptide Y and epilepsy. *Epilepsy Curr* 2003;3:53–8, doi:[10.1046/j.1531-7597.2003.030208.x](#).
66. Bedoui S, von Horsten S, Gebel A, et al. A role for neuropeptide Y (NPY) in phagocytosis: implications for innate and adaptive immunity. *Peptides* 2006;27:2037–6, doi:[10.1016/j.peptides.2006.07.029](#).
67. De la Fuente M, Del Rio A, Bedoui S. Changes with aging in the modulation by neuropeptide Y of murine peritoneal macrophage functions. *J Neuroimmunol* 2001;116:156–67, doi:[10.1016/S0165-5728\(01\)00297-1](#).
68. Dimitrijevic M, Stancijevic Vujic V, et al. Effect of neuropeptide Y on inflammatory paw edema in the rat: involvement of peripheral NPY Y1 and Y5 receptors and interaction with dipeptidyl-peptidase IV (CD26). *J Neuroimmunol* 2002;129:35–42, doi:[10.1016/S0165-5728\(02\)00173-X](#).
69. von Horsten S, Nave H, Ballof J, et al. Centrally applied NPY mimics immunoactivation induced by non-analgesic doses of met-enkephalin. *Neuroreport* 1998;9:3881–5, doi:[10.1097/00001756-199812010-00021](#).
70. von Horsten S, Ballof J, Helfritz F, et al. Modulation of innate immune functions by intracerebroventricularly applied neuropeptide Y: dose and time dependent effects. *Life Sci* 1998;63:909–22, doi:[10.1016/S0024-3205\(98\)00349-X](#).
71. Lassegue B, Clempus RE. Vascular NAD(P)H oxidases: specific features, expression, and regulation. *Am J Physiol Regul Integr Comp Physiol* 2003;285:R277–97.
72. Schallreuter KU, Elvåry S, Gibbons NC, et al. Activation/deactivation of acetylcholinesterase by H₂O₂: more evidence for oxidative stress in vitiligo. *Biochem Biophys Res Commun* 2004;315:502–8, doi:[10.1016/j.bbrc.2004.01.082](#).
73. Schallreuter KU, Vignale R, Berger J. Low catalase levels in the epidermis of patients with vitiligo. *J Invest Dermatol* 1991;97:1081–5, doi:[10.1111/j.1523-1747.ep12492612](#).
74. Kannan L. Free radical theory of autoimmunity. *Theor Biol Med Model* 2006;3:22, doi:[10.1186/1742-4682-3-22](#).
75. De la Fuente M, Bernaez I, Del Rio M, et al. Stimulation of murine peritoneal macrophage functions by neuropeptide Y and peptide YY. Involvement of protein kinase C. *Immunology* 1993;80:232–6.
76. Song C, Leonard BE. Comparison between the effects of sigma receptor ligand JO 1784 and neuropeptide Y on immune functions. *Eur J Pharmacol* 1998;345:79–87, doi:[10.1016/S0014-2999\(97\)01592-6](#).
77. Levite M. Neuropeptides, by direct interaction with T cells, induce cytokine secretion and break the commitment to a distinct T helper phenotype. *Proc Natl Acad Sci U S A* 1998;95:12544–9, doi:[10.1073/pnas.95.21.12544](#).
78. Levite M, Cahalon L, Hershkovitz R, et al. Neuropeptides, via specific receptors, regulate T cell adhesion to fibronectin. *J Immunol* 1998;160:993–1000.
79. Birol A, Kisa U, Kurtipek GS, et al. Increased tumor necrosis factor alpha (TNF-alpha) and interleukin 1 alpha (IL1-alpha) levels in the lesional skin of patients with nonsegmental vitiligo. *Int J Dermatol* 2006;45:992–3, doi:[10.1111/j.1365-4632.2006.02744.x](#).
80. Tu CX, Gu JS, Lin XR. Increased interleukin-6 and granulocyte-macrophage colony stimulating factor levels in the sera of patients with non-segmental vitiligo. *J Dermatol Sci* 2003;31:73–8, doi:[10.1016/S0923-1811\(02\)00151-2](#).
81. Bedoui S, Kuhlmann S, Nave H, et al. Differential effects of neuropeptide Y (NPY) on leukocyte subsets in the blood: mobilization of B-1-like B-lymphocytes and activated monocytes. *J Neuroimmunol* 2001;117:125–32, doi:[10.1016/S0165-5728\(01\)00328-9](#).
82. van den Boorn JG, Konijnenberg D, Dellemijn TA, et al. Autoimmune destruction of skin melanocytes by perilesional T cells from vitiligo patients. *J Invest Dermatol* 2009;129:2220–32, doi:[10.1038/jid.2009.32](#).
83. Wankowicz-Kalinska A, van den Wijngaard RM, Tigges BJ, et al. Immunopolarization of CD4+ and CD8+ T cells to type-1-like is associated with melanocyte loss in human vitiligo. *Lab Invest* 2003;83:683–95.
84. Panuncio AL, Vignale R. Ultrastructural studies in stable vitiligo. *Am J Dermatopathol* 2003;25:16–20, doi:[10.1097/00000372-200302000-00004](#).
85. Prignano F, Ricceri F, Bianchi B, et al. Dendritic cells: ultrastructural and immunophenotypical changes upon NB-UVB in vitiligo skin. *Arch Dermatol Res* 2011;303:231–8, doi:[10.1007/s00403-010-1109-5](#).
86. Kroll TM, Bommiasamy H, Boissy RE, et al. 4-Tertiary butyl phenol exposure sensitizes human melanocytes to dendritic cell-mediated

- killing: relevance to vitiligo. *J Invest Dermatol* 2005;124:798–806, doi:[10.1111/j.0022-202X.2005.23653.x](https://doi.org/10.1111/j.0022-202X.2005.23653.x).
87. Bedoui S, Lechner S, Gebhardt T, et al. NPY modulates epinephrine-induced leukocytosis via Y-1 and Y-5 receptor activation in vivo: sympathetic co-transmission during leukocyte mobilization. *J Neuroimmunol* 2002;132:25–33, doi:[10.1016/S0165-5728\(02\)00278-3](https://doi.org/10.1016/S0165-5728(02)00278-3).
88. Sung CP, Arleth AJ, Feuerstein GZ. Neuropeptide Y upregulates the adhesiveness of human endothelial cells for leukocytes. *Circ Res* 1991;68:314–8.
89. Scholzen T, Armstrong CA, Bunnett NW, et al. Neuropeptides in the skin: interactions between the neuroendocrine and the skin immune systems. *Exp Dermatol* 1998;7:81–96, doi:[10.1111/j.1600-0625.1998.tb00307.x](https://doi.org/10.1111/j.1600-0625.1998.tb00307.x).
90. Appelgren A, Appelgren B, Kopp S, et al. Neuropeptides in the arthritic TMJ and symptoms and signs from the stomatognathic system with special consideration to rheumatoid arthritis. *J Orofac Pain* 1995;9:215–25.
91. Baraniuk JN. Neurogenic mechanisms in rhinosinusitis. *Curr Allergy Asthma Rep* 2001;1:252–61, doi:[10.1007/s11882-001-0016-4](https://doi.org/10.1007/s11882-001-0016-4).
92. Pietrobon D. Migraine: new molecular mechanisms. *Neuroscientist* 2005;11:373–86, doi:[10.1177/1073858405273554](https://doi.org/10.1177/1073858405273554).
93. Misery L. Skin, immunity and the nervous system. *Br J Dermatol* 1997;137:843–50, doi:[10.1111/j.1365-2133.1997.tb01542.x](https://doi.org/10.1111/j.1365-2133.1997.tb01542.x).
94. Levite M, Chowdhury Y. Nerve-driven immunity: neuropeptides regulate cytokine secretion of T cells and intestinal epithelial cells in a direct, powerful and contextual manner. *Ann Oncol* 2001;12 Suppl 2:S19–25, doi:[10.1093/annonc/12.suppl_2.S19](https://doi.org/10.1093/annonc/12.suppl_2.S19).
95. Vause CV, Durham PL. Calcitonin gene-related peptide differentially regulates gene and protein expression in trigeminal ganglion cells: findings from array analysis. *Neurosci Lett* 2010;473:163–7, doi:[10.1016/j.neulet.2010.01.074](https://doi.org/10.1016/j.neulet.2010.01.074).
96. Dallos A, Kiss M, Polyanka H, et al. Effects of the neuropeptides substance P, calcitonin gene-related peptide, vasoactive intestinal polypeptide and galanin on the production of nerve growth factor and inflammatory cytokines in cultured human keratinocytes. *Neuropeptides* 2006;40:251–6, doi:[10.1016/j.npep.2006.06.002](https://doi.org/10.1016/j.npep.2006.06.002).
97. Hosoi J, Murphy GF, Egan CA, et al. Regulation of Langerhans cell function by nerves containing calcitonin gene-related peptide. *Nature* 1993;363:159–63, doi:[10.1038/363159a0](https://doi.org/10.1038/363159a0).
98. Ding W, Stohl LL, Nagayama T, et al. Calcitonin gene-related peptide biases Langerhans cells toward Th2-type immunity. *J Immunol* 2008;181:6020–6, doi:[10.1093/immk/181.12.6020](https://doi.org/10.1093/immk/181.12.6020).
99. Schallreuter KU, Mocae J, Wood JM, et al. In vivo and in vitro evidence for hydrogen peroxide (H₂O₂) accumulation in the epidermis of patients with vitiligo and its successful removal by a UVB-activated pseudocatalase. *J Invest Dermatol Symp Proc* 1999;4:91–6, doi:[10.1038/sj.jidsp.5640189](https://doi.org/10.1038/sj.jidsp.5640189).
100. Kaludercic N, Takimoto E, Nagayama T, et al. Monoamine oxidase A-mediated enhanced catabolism of norepinephrine contributes to adverse remodeling and pump failure in hearts with pressure overload. *Circ Res* 2010;106:193–202, doi:[10.1161/CIRCRESAHA.109.198366](https://doi.org/10.1161/CIRCRESAHA.109.198366).
101. Kim BJ, Jones HP. Epinephrine-primed murine bone marrow-derived dendritic cells facilitate production of IL-17A and IL-4 but not IFN-gamma by CD4⁺ T cells. *Brain Behav Immun* 2010;24:1126–36, doi:[10.1016/j.bbi.2010.05.003](https://doi.org/10.1016/j.bbi.2010.05.003).
102. Bai H, Cheng J, Gao X, et al. IL-17/Th17 promotes type 1 T cell immunity against pulmonary intracellular bacterial infection through modulating dendritic cell function. *J Immunol* 2009;183:5886–95, doi:[10.4049/jimmunol.0901584](https://doi.org/10.4049/jimmunol.0901584).
103. Fujino S, Andou A, Bamba S, et al. Increased expression of interleukin 17 in inflammatory bowel disease. *Gut* 2003;52:65–70, doi:[10.1136/gut.52.1.65](https://doi.org/10.1136/gut.52.1.65).
104. Nakae S, Nambu A, Sudo K, et al. Suppression of immune induction of collagen-induced arthritis in IL-17-deficient mice. *J Immunol* 2003;171:6173–7.
105. Seiderbrand J, Elborn I, Diegelmann J, et al. Role of the novel Th17 cytokine IL-17F in inflammatory bowel disease (IBD): upregulation of colonic IL-17F expression in active Crohn's disease and analysis of the IL17F p.His161Arg polymorphism in IBD. *Inflamm Bowel Dis* 2008;14:437–45, doi:[10.1002/ibd.20339](https://doi.org/10.1002/ibd.20339).
106. Vokonas H, Hirose K, Maezawa Y, et al. IL-23 and Th17 cells enhance Th2-cell-mediated eosinophilic airway inflammation in mice. *Am J Respir Crit Care Med* 2008;178:1023–32, doi:[10.1164/rccm.200801-086OC](https://doi.org/10.1164/rccm.200801-086OC).
107. Zhang Z, Zheng M, Bindas J, et al. Critical role of IL-17 receptor signaling in acute TNBS-induced colitis. *Inflamm Bowel Dis* 2006;12:382–8, doi:[10.1097/01.MIB.0000218764.06959.91](https://doi.org/10.1097/01.MIB.0000218764.06959.91).
108. Moseley TA, Haudenschild DR, Rose L, et al. Interleukin-17 family and IL-17 receptors. *Cytokine Growth Factor Rev* 2003;14:155–74, doi:[10.1016/S1359-6101\(03\)00002-9](https://doi.org/10.1016/S1359-6101(03)00002-9).
109. Kolls JK, Linden A. Interleukin-17 family members and inflammation. *Immunity* 2004;21:467–76, doi:[10.1016/j.immuni.2004.08.018](https://doi.org/10.1016/j.immuni.2004.08.018).
110. Sutton C, Brereton C, Keogh B, et al. A crucial role for interleukin (IL)-1 in the induction of IL-17-producing T cells that mediate autoimmune encephalomyelitis. *J Exp Med* 2006;203:1685–91, doi:[10.1084/jem.20060285](https://doi.org/10.1084/jem.20060285).
111. Kotake S, Udagawa N, Takahashi N, et al. IL-17 in synovial fluids from patients with rheumatoid arthritis is a potent stimulator of osteoclastogenesis. *J Clin Invest* 1999;103:1345–52, doi:[10.1172/JCI5703](https://doi.org/10.1172/JCI5703).
112. Bassiouny DA, Shaker O. Role of interleukin-17 in the pathogenesis of vitiligo. *Clin Exp Dermatol* 2011;36:292–7, doi:[10.1111/j.1365-2230.2010.03972.x](https://doi.org/10.1111/j.1365-2230.2010.03972.x).
113. Dimitrov S, Lange T, Born J. Selective mobilization of cytotoxic leukocytes by epinephrine. *J Immunol* 2010;184:503–11, doi:[10.4049/jimmunol.0902189](https://doi.org/10.4049/jimmunol.0902189).
114. Basak PY, Adiloglu AK, Koc IG, et al. Evaluation of activatory and inhibitory natural killer cell receptors in non-segmental vitiligo: a flow cytometric study. *J Eur Acad Dermatol Venereol* 2008;22:970–6, doi:[10.1111/j.1468-3083.2008.02681.x](https://doi.org/10.1111/j.1468-3083.2008.02681.x).
115. Yanagawa Y, Matsumoto M, Togashi H. Enhanced dendritic cell antigen uptake via alpha2 adrenoceptor-mediated PI3K activation following brief exposure to noradrenaline. *J Immunol* 2010;185:5762–8, doi:[10.4049/jimmunol.1001899](https://doi.org/10.4049/jimmunol.1001899).
116. Watanabe M, Tomiyama-Miyaji C, Kainuma E, et al. Role of alpha-adrenergic stimulus in stress-induced modulation of body temperature, blood glucose and innate immunity. *Immunol Lett* 2008;115:43–9, doi:[10.1016/j.imlet.2007.09.010](https://doi.org/10.1016/j.imlet.2007.09.010).
117. Lechin F, van der Dijs B, Lechin AE, et al. Natural killer cells activity and neuroimmunological treatment of cancer. *Clin Cancer Res* 2004;10:8120, author reply 8120–1, doi:[10.1158/1078-0432.CCR-04-1508](https://doi.org/10.1158/1078-0432.CCR-04-1508).

118. Chin JH, Okazaki M, Hu ZW, et al. Activation of heat shock protein (hsp)70 and proto-oncogene expression by alpha1 adrenergic agonist in rat aorta with age. *J Clin Invest* 1996;97:2316–23, doi:[10.1172/JCI118674](https://doi.org/10.1172/JCI118674).
119. Heneka MT, Gavriluk V, Landreth GE, et al. Noradrenergic depletion increases inflammatory responses in brain: effects on IkappaB and HSP70 expression. *J Neurochem* 2003;85:387–98, doi:[10.1046/j.1471-4159.2003.01694.x](https://doi.org/10.1046/j.1471-4159.2003.01694.x).
120. Meng X, Brown JM, Ao L, et al. Norepinephrine induces cardiac heat shock protein 70 and delayed cardioprotection in the rat through alpha 1 adrenoceptors. *Cardiovasc Res* 1996;32:374–83, doi:[10.1016/0008-6363\(96\)00078-8](https://doi.org/10.1016/0008-6363(96)00078-8).
121. Johnson JD, Campisi J, Sharkey CM, et al. Adrenergic receptors mediate stress-induced elevations in extracellular Hsp72. *J Appl Physiol* 2005;99:1789–95, doi:[10.1152/japplphysiol.00390.2005](https://doi.org/10.1152/japplphysiol.00390.2005).
122. Multhoff G, Mizzen L, Winchester CC, et al. Heat shock protein 70 (Hsp70) stimulates proliferation and cytolytic activity of natural killer cells. *Exp Hematol* 1999;27:1627–36, doi:[10.1016/S0301-472X\(99\)00104-6](https://doi.org/10.1016/S0301-472X(99)00104-6).
123. Multhoff G, Botzler C, Wiesnet M, et al. A stress-inducible 72-kDa heat-shock protein (HSP72) is expressed on the surface of human tumor cells, but not on normal cells. *Int J Cancer* 1995;60:272–9, doi:[10.1002/ijc.2910610222](https://doi.org/10.1002/ijc.2910610222).
124. Barreto A, Gonzalez JM, Kabingu E, et al. Stress-induced release of HSC70 from human tumors. *Cell Immunol* 2003;222:97–104, doi:[10.1016/S0008-8749\(03\)00115-1](https://doi.org/10.1016/S0008-8749(03)00115-1).
125. Moseley P. Stress proteins and the immune response. *Immunopharmacology* 2000;48:299–302, doi:[10.1016/S0162-3109\(00\)00227-7](https://doi.org/10.1016/S0162-3109(00)00227-7).
126. Moseley PL. Heat shock proteins and the inflammatory response. *Ann N Y Acad Sci* 1998;856:206–13, doi:[10.1111/j.1749-6632.1998.tb08327.x](https://doi.org/10.1111/j.1749-6632.1998.tb08327.x).
127. Srivastava P. Interaction of heat shock proteins with peptides and antigen presenting cells: chaperoning of the innate and adaptive immune responses. *Annu Rev Immunol* 2002;20:395–425, doi:[10.1146/annurev.immunol.20.100301.064801](https://doi.org/10.1146/annurev.immunol.20.100301.064801).
128. Lehner T, Wang Y, Whittall T, et al. Functional domains of HSP70 stimulate generation of cytokines and chemokines, maturation of dendritic cells and adjuvanticity. *Biochem Soc Trans* 2004;32:629–32, doi:[10.1042/BST0320629](https://doi.org/10.1042/BST0320629).
129. Asea A, Kraeft SK, Kurt-Jones EA, et al. HSP70 stimulates cytokine production through a CD14-dependent pathway, demonstrating its dual roles as a chaperone and cytokine. *Nat Med* 2000;6:435–42, doi:[10.1038/74697](https://doi.org/10.1038/74697).
130. Campisi J, Fleshner M. Role of extracellular HSP72 in acute stress-induced potentiation of innate immunity in active rats. *J Appl Physiol* 2003;94:512–52.
131. Campisi J, Leem TH, Fleshner M. Stress-induced extracellular Hsp72 is a functionally significant danger signal to the immune system. *Cell Stress Chaperones* 2003;8:272–86, doi:[10.1379/1466-1268\(2003\)008<0272:SEHIAF>2.0.CO;2](https://doi.org/10.1379/1466-1268(2003)008<0272:SEHIAF>2.0.CO;2).
132. Multhoff G, Botzler C, Jennen L, et al. Heat shock protein 72 on tumor cells: a recognition structure for natural killer cells. *J Immunol* 1997;158:4341–50.
133. Niland B, Miklossy G, Banki K, et al. Cleavage of transaldolase by granzyme B causes the loss of enzymatic activity with retention of antigenicity for multiple sclerosis patients. *J Immunol* 2010;184:4025–32, doi:[10.4049/jimmunol.0804174](https://doi.org/10.4049/jimmunol.0804174).
134. Le Poole IC, Luiten RM. Autoimmune etiology of generalized vitiligo. *Curr Dir Autoimmun* 2008;10:227–43, doi:[10.1159/000131485](https://doi.org/10.1159/000131485).
135. Denman CJ, McCracken J, Krishnan V, et al. HSP70i accelerates depigmentation in a mouse model of autoimmune vitiligo. *J Invest Dermatol* 2008;128:2041–8, doi:[10.1038/jid.2008.45](https://doi.org/10.1038/jid.2008.45).
136. Yuan ZQ, Zhang Y, Li XL, et al. HSP70 protects intestinal epithelial cells from hypoxia/reoxygenation injury via a mechanism that involves the mitochondrial pathways. *Eur J Pharmacol* 2010;643:202–8, doi:[10.1016/j.ejphar.2010.06.068](https://doi.org/10.1016/j.ejphar.2010.06.068).
137. Kawana K, Miyamoto Y, Tanonaka K, et al. Cytoprotective mechanism of heat shock protein 70 against hypoxia/reoxygenation injury. *J Mol Cell Cardiol* 2000;32:2229–37, doi:[10.1006/jmcc.2000.1270](https://doi.org/10.1006/jmcc.2000.1270).
138. Magalhães J, Ascensao A, Soares JM, et al. Acute and severe hypobaric hypoxia increases oxidative stress and impairs mitochondrial function in mouse skeletal muscle. *J Appl Physiol* 2005;99:1247–53, doi:[10.1152/japplphysiol.01324.2004](https://doi.org/10.1152/japplphysiol.01324.2004).
139. Tokyol C, Karaorman G, Bastug M. Effects of acute and adaptive hypoxia on heat shock protein expression in hepatic tissue. *High Alt Med Biol* 2005;6:247–55, doi:[10.1089/ham.2005.6.247](https://doi.org/10.1089/ham.2005.6.247).
140. Renis M, Cardile V, Grasso S, et al. Switching off HSP70 and i-NOS to study their role in normal and H2O2-stressed human fibroblasts. *Life Sci* 2003;74:757–69, doi:[10.1016/j.lfs.2003.07.016](https://doi.org/10.1016/j.lfs.2003.07.016).
141. Calabrese V, Scapagnini G, Catalano C, et al. Regulation of heat shock protein synthesis in human skin fibroblasts in response to oxidative stress: role of vitamin E. *Int J Tissue React* 2001;23:127–35.
142. Scarparo AC, Visconti MA, de Oliveira AR, et al. Adrenoceptors in normal and malignant human melanocytes. *Arch Dermatol Res* 2000;292:265–7, doi:[10.1007/s004030050485](https://doi.org/10.1007/s004030050485).
143. Johnson EM Jr, Gorin PD, Brandeis LD, et al. Dorsal root ganglion neurons are destroyed by exposure in utero to maternal antibody to nerve growth factor. *Science* 1980;210:916–8, doi:[10.1126/science.7192014](https://doi.org/10.1126/science.7192014).
144. Levi-Montalcini R, Angeletti PU. Nerve growth factor. *Physiol Rev* 1968;48:534–69.
145. Meakin SO, Shooter EM. The nerve growth factor family of receptors. *Trends Neurosci* 1992;15:323–31, doi:[10.1016/0166-2236\(92\)90047-C](https://doi.org/10.1016/0166-2236(92)90047-C).
146. Chao MV, Hempstead BL. p75 and Trk: a two-receptor system. *Trends Neurosci* 1995;18:321–6, doi:[10.1016/0166-2236\(95\)93922-K](https://doi.org/10.1016/0166-2236(95)93922-K).
147. Levi-Montalcini R, Skaper SD, Dal Toso R, et al. Nerve growth factor: from neurotrophin to neurokin. *Trends Neurosci* 1996;19:514–20, doi:[10.1016/S0166-2236\(96\)10058-8](https://doi.org/10.1016/S0166-2236(96)10058-8).
148. Bischoff SC, Dahinden CA. Effect of nerve growth factor on the release of inflammatory mediators by mature human basophils. *Blood* 1992;79:2662–9.
149. Purcell WM, Atterwill CK. Mast cells in neuroimmune function: neurotoxicological and neuropharmacological perspectives. *Neurochem Res* 1995;20:521–32, doi:[10.1007/BF01694534](https://doi.org/10.1007/BF01694534).
150. Torcia M, Bracci-Laudiero L, Lucibello M, et al. Nerve growth factor is an autocrine survival factor for memory B lymphocytes. *Cell* 1996;85:345–56, doi:[10.1016/S0092-8674\(00\)81113-7](https://doi.org/10.1016/S0092-8674(00)81113-7).
151. Torii H, Yan Z, Hosoi J, et al. Expression of neurotrophic factors and neuropeptide receptors by Langerhans cells and the Langerhans cell-like cell line XS2: further support for a functional relationship between Langerhans cells and epidermal

- nerves. *J Invest Dermatol* 1997;109:586–91, doi:[10.1111/1523-1747.ep12337516](https://doi.org/10.1111/1523-1747.ep12337516).
152. Lambiasi A, Bracci-Laudiero L, Bonini S, et al. Human CD4+ T cell clones produce and release nerve growth factor and express high-affinity nerve growth factor receptors. *J Allergy Clin Immunol* 1997;100:408–14, doi:[10.1016/S0091-6749\(97\)70256-2](https://doi.org/10.1016/S0091-6749(97)70256-2).
153. Garaci E, Caroleo MC, Aloe L, et al. Nerve growth factor is an autocrine factor essential for the survival of macrophages infected with HIV. *Proc Natl Acad Sci U S A* 1999;96:14013–8, doi:[10.1073/pnas.96.24.14013](https://doi.org/10.1073/pnas.96.24.14013).
154. Thorpe LW, Perez-Polo JR. The influence of nerve growth factor on the in vitro proliferative response of rat spleen lymphocytes. *J Neurosci Res* 1987;18:134–9, doi:[10.1002/jnr.490180120](https://doi.org/10.1002/jnr.490180120).
155. Otten U, Ehrhard P, Peck R. Nerve growth factor induces growth and differentiation of human B lymphocytes. *Proc Natl Acad Sci U S A* 1989;86:10059–63, doi:[10.1073/pnas.86.24.10059](https://doi.org/10.1073/pnas.86.24.10059).
156. Kimata H, Yoshida A, Ishioka C, et al. Nerve growth factor specifically induces human IgG4 production. *Eur J Immunol* 1991;21:137–41, doi:[10.1002/eji.1830210121](https://doi.org/10.1002/eji.1830210121).
157. Brodie C, Gelfand EW. Functional nerve growth factor receptors on human B lymphocytes. Interaction with IL-2. *J Immunol* 1992;148:3492–7.
158. Kemp EH, Emhemad S, Akhtar S, et al. Autoantibodies against tyrosine hydroxylase in patients with non-segmental (generalised) vitiligo. *Exp Dermatol* 2011;20:35–40, doi:[10.1111/j.1600-0625.2010.01181.x](https://doi.org/10.1111/j.1600-0625.2010.01181.x).
159. Kemp EH, Gawkrödger DJ, MacNeil S, et al. Detection of tyrosinase autoantibodies in patients with vitiligo using 35S-labeled recombinant human tyrosinase in a radioimmunoassay. *J Invest Dermatol* 1997;109:69–73, doi:[10.1111/j.1523-1747.ep12276556](https://doi.org/10.1111/j.1523-1747.ep12276556).
160. Kemp EH, Gawkrödger DJ, Watson PF, et al. Immunoprecipitation of melanocyte-specific autoantigens with vitiligo sera: evidence for cross-reactive autoantibodies to tyrosinase and tyrosinase-related protein-2 (TRP-2). *Clin Exp Immunol* 1997;109:495–500, doi:[10.1046/j.1365-2249.1997.4781381.x](https://doi.org/10.1046/j.1365-2249.1997.4781381.x).
161. Kemp EH, Gawkrödger DJ, Watson PF, et al. Autoantibodies to human melanocyte-specific protein mel17 in the sera of vitiligo patients: a sensitive and quantitative radioimmunoassay (RIA). *Clin Exp Immunol* 1996;107:333–8, doi:[10.1046/j.1365-2249.1998.00746.x](https://doi.org/10.1046/j.1365-2249.1998.00746.x).
162. Aloe L, Bracci-Laudiero L, Bonini S, et al. The expanding role of nerve growth factor: from neurotrophic activity to immunologic diseases. *Allergy* 1997;52:883–94, doi:[10.1111/j.1398-9995.1997.tb01247.x](https://doi.org/10.1111/j.1398-9995.1997.tb01247.x).
163. Horigome K, Burchick EL, Johnson EM, Jr. Effects of nerve growth factor on rat peritoneal mast cells. Survival promotion and immediate-early gene induction. *J Biol Chem* 1994;269:2695–702.
164. Solomon A, Aloe L, Pe'er J, et al. Nerve growth factor is preformed in and activates human peripheral blood eosinophils. *J Allergy Clin Immunol* 1998;102:454–60, doi:[10.1016/S0091-6749\(98\)70135-6](https://doi.org/10.1016/S0091-6749(98)70135-6).
165. Gawkrödger DJ, Ormerod AD, Shaw L, et al. Vitiligo: concise evidence based guidelines on diagnosis and management. *Postgrad Med J* 2010;86:466–71, doi:[10.1136/pgmj.2009.093278](https://doi.org/10.1136/pgmj.2009.093278).
166. Hossani-Madani AR, Halder RM. Topical treatment and combination approaches for vitiligo: new insights, new developments. *G Ital Dermatol Venereol* 2010;145:57–78.
167. Taher ZA, Lauzon G, Maguiness S, et al. Analysis of interleukin-10 levels in lesions of vitiligo following treatment with topical tacrolimus. *Br J Dermatol* 2009;161:654–9, doi:[10.1111/j.1365-2133.2009.09217.x](https://doi.org/10.1111/j.1365-2133.2009.09217.x).
168. Grimbaldston N, Nakae S, Kalesnikoff J, et al. Mast cell-derived interleukin 10 limits skin pathology in contact dermatitis and chronic irradiation with ultraviolet B. *Nat Immunol* 2007;8:1095–104, doi:[10.1038/ni1503](https://doi.org/10.1038/ni1503).
169. Braat H, Rottiers P, Hommes DW, et al. A phase I trial with transgenic bacteria expressing interleukin-10 in Crohn's disease. *Clin Gastroenterol Hepatol* 2006;4:754–9, doi:[10.1016/j.cgh.2006.03.020](https://doi.org/10.1016/j.cgh.2006.03.020).
170. Krdžević Fijan S, Furnsinn-Friedl AM, Honigsmann H, et al. Oral dexamethasone pulse treatment for vitiligo. *J Am Acad Dermatol* 2001;44:814–7, doi:[10.1067/mjd.2001.113475](https://doi.org/10.1067/mjd.2001.113475).
171. Bai A, Lu N, Guo Y, et al. Modulation of inflammatory response via alpha2-adrenoceptor blockade in acute murine colitis. *Clin Exp Immunol* 2009;156:353–62, doi:[10.1111/j.1365-2249.2009.03894.x](https://doi.org/10.1111/j.1365-2249.2009.03894.x).
172. Harmon RJ, Riggs PD. Clonidine for posttraumatic stress disorder in preschool children. *J Am Acad Child Adolesc Psychiatry* 1996;35:1247–9, doi:[10.1097/00004583-199609000-00022](https://doi.org/10.1097/00004583-199609000-00022).
173. Kinzie JD, Leung P. Clonidine in Cambodian patients with posttraumatic stress disorder. *J Nerv Ment Dis* 1989;177:546–50, doi:[10.1097/00005053-198909000-00005](https://doi.org/10.1097/00005053-198909000-00005).
174. Stander S, Stander H, Seeliger S, et al. Topical pimecrolimus and tacrolimus transiently induce neuropeptide release and mast cell degranulation in murine skin. *Br J Dermatol* 2007;156:1020–6, doi:[10.1111/j.1365-2133.2007.07813.x](https://doi.org/10.1111/j.1365-2133.2007.07813.x).
175. Gupta MA, Guptat AK. The use of antidepressant drugs in dermatology. *J Eur Acad Dermatol Venereol* 2001;15:512–8, doi:[10.1046/j.1468-3083.2001.00278.x](https://doi.org/10.1046/j.1468-3083.2001.00278.x).
176. Papadopoulos L, Bor R, Legg C. Coping with the disfiguring effects of vitiligo: a preliminary investigation into the effects of cognitive-behavioural therapy. *Br J Med Psychol* 1999;72(Pt 3):385–96, doi:[10.1348/000711299160077](https://doi.org/10.1348/000711299160077).
177. Jin Y, Birlea SA, Fain PR, et al. Genetic variations in NALP1 are associated with generalized vitiligo in a Romanian population. *J Invest Dermatol* 2007;127:2558–62, doi:[10.1038/sj.jid.5700953](https://doi.org/10.1038/sj.jid.5700953).
178. Jin Y, Mailloux CM, Gowan K, et al. NALP1 in vitiligo-associated multiple autoimmune disease. *N Engl J Med* 2007;356:1216–25, doi:[10.1056/NEJMoa061592](https://doi.org/10.1056/NEJMoa061592).
179. Jin Y, Birlea SA, Fain PR, et al. Genome-wide analysis identifies a quantitative trait locus in the MHC class II region associated with generalized vitiligo age of onset. *J Invest Dermatol* 2011;131:1308–12.
180. Birlea SA, Jin Y, Bennett DC, et al. Comprehensive association analysis of candidate genes for generalized vitiligo supports XBP1, FOXP3, and TSLP. *J Invest Dermatol* 2011;131:371–81, doi:[10.1038/jid.2010.337](https://doi.org/10.1038/jid.2010.337).
181. Spritz RA. The genetics of generalized vitiligo: autoimmune pathways and an inverse relationship with malignant melanoma. *Genome Med* 2010;2:78, doi:[10.1186/gm199](https://doi.org/10.1186/gm199).
182. Spritz RA. Shared genetic relationships underlying generalized vitiligo and autoimmune thyroid disease. *Thyroid* 2010;20:745–54, doi:[10.1089/thy.2010.1643](https://doi.org/10.1089/thy.2010.1643).
183. Jin Y, Birlea SA, Fain PR, et al. Common variants in FOXP1 are associated with generalized vitiligo. *Nat Genet* 2010;42:576–8, doi:[10.1038/ng.602](https://doi.org/10.1038/ng.602).

184. Quan C, Ren YQ, Xiang LH, et al. Genome-wide association study for vitiligo identifies susceptibility loci at 6q27 and the MHC. *Nat Genet* 2010;42:614–8, doi:[10.1038/ng.603](https://doi.org/10.1038/ng.603).
185. Ren Y, Yang S, Xu S, et al. Genetic variation of promoter sequence modulates XBP1 expression and genetic risk for vitiligo. *PLoS Genet* 2009;5:e1000523, doi:[10.1371/journal.pgen.1000523](https://doi.org/10.1371/journal.pgen.1000523).
186. Liang Y, Yang S, Zhou Y, et al. Evidence for two susceptibility loci on chromosomes 22q12 and 6p21-p22 in Chinese generalized vitiligo families. *J Invest Dermatol* 2007;127:2552–7, doi:[10.1038/sj.jid.5700904](https://doi.org/10.1038/sj.jid.5700904).
187. Xu S, Zhou Y, Yang S, et al. Platelet-derived growth factor receptor alpha gene mutations in vitiligo vulgaris. *Acta Derm Venereol* 2010;90:131–5, doi:[10.2340/00015555-0820](https://doi.org/10.2340/00015555-0820).

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